

Singlet Oxygen Chemistry in Water: A Porous Vycor Glass-Supported Photosensitizer

David Aebisher,[†] Nikolay S. Azar,[†] Matibur Zamadar,[†] Naveen Gandra,[‡] Harry D. Gafney,[§] Ruomei Gao,^{*,‡} and Alexander Greer^{*,†}

Department of Chemistry and Graduate Center, City University of New York, Brooklyn College, Brooklyn, New York 11210, Department of Chemistry, Jackson State University, Jackson, Mississippi 39217, and Department of Chemistry and Biochemistry & Graduate Center, City University of New York, Queens College, Flushing, New York, 11367

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Singlet molecular oxygen [$^1\text{O}_2$ ($^1\Delta_g$)] is generated cleanly in aqueous solution upon irradiation of a heterogeneous complex, *meso*-tetra(*N*-methyl-4-pyridyl)porphine (**1**) adsorbed onto porous Vycor glass (PVG). The cationic photosensitizer **1** tightly binds onto PVG and gives a stable material, which does not dissociate **1** into the surrounding aqueous phase. The production of $^1\text{O}_2$ was measured by monitoring the time-resolved $^1\text{O}_2$ ($^1\Delta_g$) phosphorescence at 1270 nm. Indirect analysis of $^1\text{O}_2$ generation was also carried out with the photooxidation of *trans*-2-methyl-2-pentenoate anion, which afforded the corresponding hydroperoxide. Sensitizer-**1**-impregnated PVG gives rise to a new singlet oxygen generator but more importantly provides a heterogeneous system for use in water.

1. Introduction

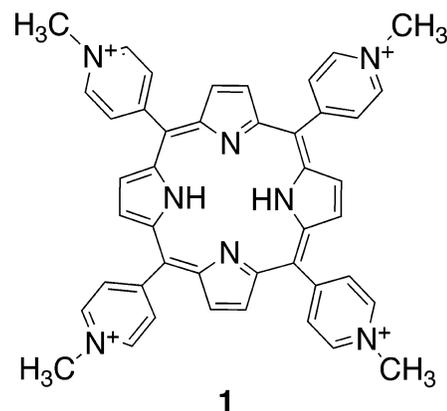
Singlet oxygen [$^1\text{O}_2$ ($^1\Delta_g$)] has a lifetime of microseconds to milliseconds and can diffuse a distance from where it was generated.^{1,2} Remarkably, $^1\text{O}_2$ can diffuse across a cell membrane^{3,4} and even serve as a signaling molecule.^{5–7} The first report of $^1\text{O}_2$ as a diffusible intermediate came from Kautsky and de Bruijn in 1931,⁸ in which tryptaflavine (a sensitizer) and leucomalachite green (an oxygen-acceptor compound) were adsorbed separately on silica gel beads (Scheme 1). Upon photolysis in the presence of O_2 , $^1\text{O}_2$ was generated on a SiO_2 bead and diffused to another (separate) SiO_2 bead where it was trapped by leucomalachite green.^{8–10} Kautsky's "through space" experiment was similar in many respects to the Paneth/Hofeditz lead mirror experiments (thermal decomposition of PbEt_4),¹¹ which provided evidence for free ethyl radicals in the gas phase.^{12–14} Other researchers have since conducted similar heterogeneous $^1\text{O}_2$ studies. The first heterogeneous photosensitizer (covalently bound) was polymer Rose Bengal synthesized by Neckers et al. in 1972.^{15–17} Polymer Rose Bengal was based on Merrifield beads with its origins traced directly to solid-phase peptide synthetic chemistry.

Silica gel, polymers, alumina, and zeolites have been used as solid supports for $^1\text{O}_2$ photosensitizers.¹⁸ However, these solid supports^{18a–c} are often used to generate $^1\text{O}_2$ in the presence of organic solvents, or in the absence of solvent altogether.

Additional studies could be directed toward $^1\text{O}_2$ chemistry in water, and the search for alternative solid supports. We sought a system that does not require covalent attachment of the sensitizer to the solid but nonetheless binds the two together tightly. Because porous Vycor glass (PVG) develops a negative ζ potential in water (indicating an anionic surface), cations bind tightly.^{19,20} Neutral molecules can also bind to PVG^{21–27} but

do so more weakly than cations. By taking advantage of the cation-binding ability of PVG in water,^{19,20,28–32} one can imagine a similar PVG binding of a cationic photosensitizer. Raftery et al. have used PVG as a solid support for photocatalysts, such as a TiO_2 monolayer.^{33,34} To date, no reports exist on heterogeneous $^1\text{O}_2$ photooxidations with PVG as a solid support. *Meso*-tetra(*N*-methyl-4-pyridyl)porphine (**1**) was selected in our study because it is a $^1\text{O}_2$ photosensitizer² known to photodynamically inactivate *E. coli*.³⁵ Anions tend not to bind to PVG. Thus, PVG might also yield specificity for $^1\text{O}_2$ reactions with anions rather than cations and neutral compounds in the bulk aqueous phase.

We report that cation **1** binds to the PVG anionic silanol sites to give an adsorbed complex. Singlet oxygen is photochemically generated at the solid–liquid interface, and then, $^1\text{O}_2$ diffuses into the aqueous medium. An anionic alkene is readily oxidized by $^1\text{O}_2$ in the surrounding aqueous solution.



2. Experimental Section

2.1. Materials and Instrumentation. Reagents were obtained commercially [*meso*-tetra(*N*-methyl-4-pyridyl)porphine tetratosylate, *meso*-tetra(4-sulfonatophenyl)porphine dihydro-chloride, *trans*-2-methyl-2-pentenoic acid, sodium hydroxide, magnesium

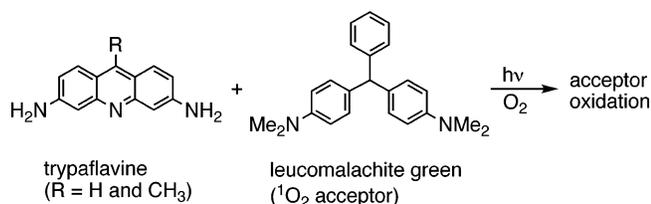
* E-mail: ruomei.gao@jsums.edu (R.G.); agreer@brooklyn.cuny.edu (A.G.).

[†] Brooklyn College.

[‡] Jackson State University.

[§] Queens College.

SCHEME 1



sulfate, triphenylphosphine, *p*-toluene sulfonic acid (TsOH), and adipic acid] and used without further purification. The solvents used (methanol, absolute ethanol, deuterium oxide-*d*₂, and chloroform-*d*₁) were of spectroscopic or equivalent grade and were used as received. Deionized water was obtained from a U.S. Filter Corporation deionization system. PVG samples (Corning 7930, pore size = 40 Å) were dried in a Fisher Scientific Isotemp muffle furnace at 500 °C and then stored in a desiccator under a vacuum (30 mmHg). Pieces of PVG were in the shape of disks (11.0 mm in diameter and 2.0 mm in thickness) or squares (1.5 cm² and 1.0–1.5 mm in thickness). Samples were irradiated with a *Q*-switched Nd:YAG laser (532 nm, 3–4 ns, 30 mJ, Polaris II-20, New Wave Research Merchantek Products). A liquid-N₂-cooled germanium photodetector (Applied Detector Corporation) was used for the determination of quantum yields of ¹O₂. The steady-state generation of ¹O₂ was conducted with a Rayonet photoreactor with Sylvania F8T5/CW 8 W bulbs that emit at ~425–650 nm. Mass spectrometry data were acquired on an Agilent Technologies 6890N GC/MS instrument with a 5973 mass selective detector (MSD) and a HP-5MS column. UV–visible spectra were collected on a Hitachi UV–vis U-2001 instrument. NMR data were collected on a Bruker DPX400 NMR instrument.

2.2. Measurements. The adsorption process was achieved by placing a 0.28 g PVG sample into 24.7 mL of a 1.0 × 10⁻⁵ M solution of **1** in deionized water. The amount of photosensitizer adsorbed onto PVG was calculated from the difference in absorbance of the solution before introduction of PVG and the absorbance of the same solution after the PVG's removal [at the λ_{max} value of **1** (422 nm)]. The number of moles of photosensitizer adsorbed per gram of PVG was calculated using eq 1

$$n_{\text{adsorbed}}/\text{g of PVG} = \{[(A_i - A_f)/A_i] \times n_i\}/\text{g of PVG} \quad (1)$$

in which *A*_i is the absorbance of the solution prior to introduction of PVG, *A*_f is the absorbance of the solution after the impregnation interval and removal of the PVG, *n*_i is the number of moles of sensitizer prior to impregnation, *n*_{adsorbed} is the number of moles adsorbed onto the PVG, and *g* of PVG is the weight of the PVG in grams.²⁹ Typically, PVG samples were loaded with 1.0 × 10⁻⁶ mol of **1**/g of PVG. Colorless PVG was converted to deep red on adsorption of **1** after 48 h. The moles of hydronium ion were calculated from the pH of the surrounding aqueous solution. The experiments were carried out at room temperature and with O₂-saturated solutions. The phosphorescence of ¹O₂ at 1270 nm was measured as previously described.³⁶ The initial ¹O₂ intensity is extrapolated to *t* = 0. The data points of the initial 3–4 ns are not used due to electronic interference signals from the detector. The intensity of the pulses at 532 nm was controlled between 20 and 30 mJ. The quantum yield for production of ¹O₂ in homogeneous solution is calculated according to eq 2.

$$\Phi_{\Delta \text{ sample}}/\Phi_{\Delta \text{ reference}} = S_{\text{sample}}/S_{\text{reference}} \quad (2)$$

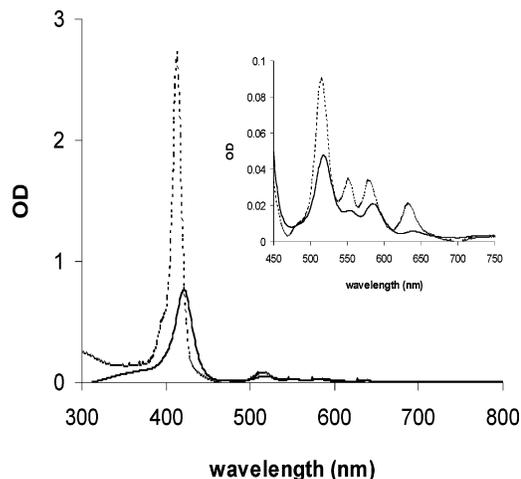


Figure 1. Absorption spectra of *meso*-tetra(4-sulfonatophenyl)porphine (dashed line) and **1** (solid line) in H₂O solution. The inset is an expanded view of the visible portion of the spectra, in which the two compounds are optically matched at 532 nm.

Here, Φ_{Δ sample} and Φ_{Δ reference} are the ¹O₂ quantum yields for samples and the reference. *Meso*-tetra(4-sulfonatophenyl)porphine was used as a reference sensitizer, in which its absorbance was optically matched with that of **1** at 532 nm (Figure 1). *S*_{sample} and *S*_{reference} represent the slopes obtained from the plot of initial intensity of ¹O₂ via the absorbance at an excitation wavelength of 532 nm for the sample and the reference, respectively.

2.3. *Meso*-tetra(*N*-methyl-4-pyridyl)porphine (1**).** The purity of the tosylate of **1** was determined to be 99% based on NMR (integration of peaks in its ¹H NMR spectrum) and greater than 95% based on HRMS: ¹H NMR (D₂O, 400 MHz) δ 1.50 (s, 12H), 4.63 (s, 12H), 6.43 (bs, 8H), 7.01 (bs, 8H), 8.78 (d, *J* = 4.6 Hz, 8H), 8.83–9.08 (bs, 8H), 9.14 (d, *J* = 4.6 Hz, 8H). ¹³C NMR (D₂O, 100 MHz) δ 155.0, 141.6, 138.7, 136.9, 130.8, 126.1, 122.3, 113.3, 46.1, 17.1. Mass spectrum (EI); *m/z* = 676.31 (100), 677.31 (48), 678.31 (12.5), 677.30 (3.0), 679.32 (1.7). HRMS calcd for (C₄₄H₃₆N₈)²⁺ 338.1522 (the base peak represents the loss of four tosylate fragments and two protons), found 338.1525. Literature: ¹H NMR (D₂O) δ 9.19 (d, 8H), 9.03 (bs, 8H), 8.71 (d, 8H), 4.77 (s, 12H) at pD 7.0.³⁷ ¹H NMR (DMSO) δ 9.47 (d, 8H), 9.18 (s, 8H), 8.97 (d, 8H), 4.72 (s, 12H).³⁸

2.4. Heterogeneous Photooxidation of *Trans*-2-methyl-2-pentenoate Anion (2**).** Compound **1** (7.5 × 10⁻⁷ mol) adsorbed onto 1.58 g of PVG was combined with 20 mL of *trans*-2-methyl-2-pentenoic acid (0.25 mmol) and NaOH (0.35 mmol) in deionized H₂O. Presumably *trans*-2-methyl-2-pentenoic acid is a stronger acid and is selectively deprotonated affording **2**, since the PVG silanol groups are reported to be p*K*_a ~ 9.³⁹ Dioxygen was bubbled into the solutions. Photooxygenation was carried out for 4 h at room temperature with the Rayonet reactor. Aliquots of the H₂O reaction were mixed with D₂O for NMR analysis. 3-Hydroperoxy-2-methylene pentanoic acid (**3**) was detected as the sole product: ¹H NMR (D₂O) δ 0.91 (t, *J* = 7.5 Hz, 3H), 1.66 (m, 2H), 4.75 (t, *J* = 6.5 Hz, 1H), 5.56 (s, 1H), 5.94 (s, 1H). ¹H NMR (CDCl₃) δ 0.97 (t, *J* = 7.4 Hz, 3H), 1.71 (m, 2H), 4.82 (t, *J* = 6.4 Hz, 1H), 6.01 (s, 1H), 6.54 (s, 1H) [note that 1.66 (in D₂O) and 1.71 (CDCl₃) represent ABX3 multiplets]. HRMS calcd for (C₆H₉O₄)(M – H⁺) 145.0506, found 145.0506. The percent yield was determined by comparison of the integrated methyl protons of **3** with the methylene protons of adipic acid (internal standard) by ¹H NMR spectroscopy. Upon addition of triphenylphosphine (0.08 M), **3** converted to 3-hydroxy-2-methylene pentanoic acid (**4**),

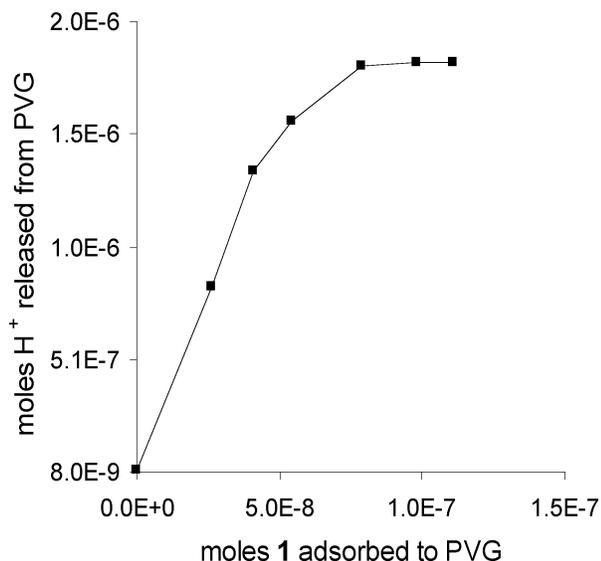


Figure 2. Plot of the moles of H⁺ ions arising from PVG silanol deprotonation calculated from the decrease in pH of the aqueous solution during the adsorption of **1** onto PVG. Each point is taken at a 30 min increment over a total of 3 h.

a known compound.⁴⁰ Our data agreed with the literature value: ⁴⁰ ¹H NMR (D₂O) δ 1.19 (t, *J* = 7.1 Hz, 3H), 1.54 (m, 2H), 4.44 (t, *J* = 6.0 Hz, 1H), 5.45 (s, 1H), 5.76 (s, 1H) (note that 1.54 represents an ABX3 multiplet).

3. Results and Discussion

3.1. Adsorption of Photosensitizer 1. In aqueous solution, cation **1** adsorbs onto PVG and forms a complex. After 48 h, we find that 1.0×10^{-6} mol of **1** adsorbed onto 1 g of PVG. Three experiments were conducted to examine aspects of the adsorption process. First, an experiment was carried out in order to determine whether a decrease in pH in the surrounding solution coincides with the adsorption of **1** onto PVG. A concurrent decrease in pH of the aqueous solution is found (plotted as the appearance of moles of H⁺ ions, Figure 2), which suggests that cation **1** exchanges onto the anionic silanol sites. The adsorption of **1** over 3 h corresponded to a pH reduction from 6.12 to 4.14. Figure 1 shows that cationic **1** replaces hydronium ions on the PVG silanol groups. The initial mole ratio of **1** adsorbed/H⁺ dissociated is 1:32 (after 30 min). After 3 h, an equilibrium is established and reveals that every mole of **1** adsorbed leads to the release of 15 mol of H⁺ from PVG. Perhaps this decrease in the ratio relates to a tendency for reprotonation of some silanol anion sites over time. Second, to determine if the counterion (TsO⁻) is coadsorbed onto the anionic surface, a 1.69 g PVG sample was placed in 25.0 mL of 1.01×10^{-5} M **1** and the aqueous phase was monitored by UV spectroscopy. Adsorption of **1** was accompanied by the appearance of 3.9 ± 0.05 mol of equivalent tosylate ion in solution ($\lambda_{\max} = 260$ nm) measured by a prior constructed calibration curve. The spectroscopic analysis suggested that less than 5% TsO⁻ counterion coadsorbs onto PVG, and that the majority of TsO⁻ remains in the surrounding aqueous solution likely because of the Coulombic interaction disfavoring association of the anion with the anionic silanol surface of PVG. Similarly, Cr³⁺, Fe³⁺, and Cu²⁺ are known to cation exchange onto PVG with less than 3% coadsorption of the Cl⁻ counterion.²⁹ Third, repeated washing with water failed to detect **1** by UV-visible absorption (the detection limit of **1** is 3.1×10^{-7} M at $\lambda_{\max} = 422$ nm). One may assume the worst case, i.e.,

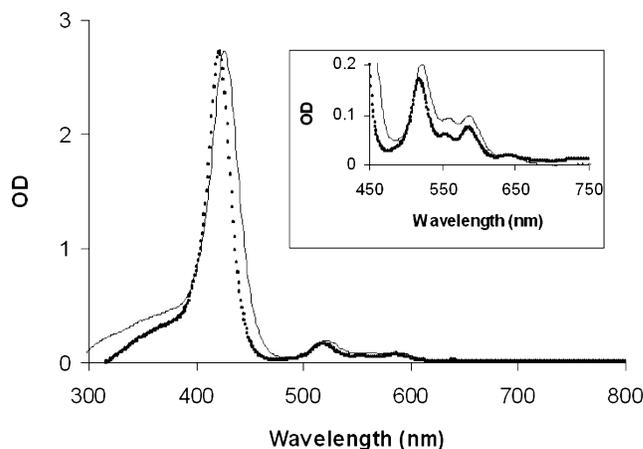


Figure 3. Normalized absorption spectra of **1-ads** and **1**: (1) the solid line is of **1-ads** in air, in which uncoated PVG was used as a blank, and (2) the dashed line is an H₂O solution of **1**, in which H₂O was used as a blank. The inset is an expanded view of the visible portion of the spectra.

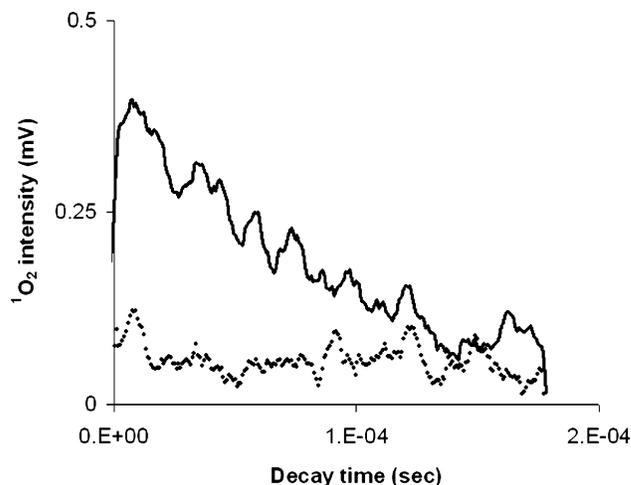


Figure 4. Singlet oxygen phosphorescence (1270 nm) decay from **1-ads** carried out in H₂O (dots) and D₂O (solid line). The above traces have been corrected, in which an uncoated PVG sample served as a blank and was subtracted from the background.

desorption below the detection limit of $\sim 3 \times 10^{-7}$ M **1** of our UV instrument. However, this conclusion is probably incorrect because a control experiment showed that 3×10^{-7} M **1** does not lead to the ¹O₂ chemistry in sections 3.3 and 3.4. The experimental data lead us to conclude that the adsorbed **1** remains attached to PVG throughout the sensitization and photochemical processes. Thus, the PVG-attached sensitizer is referred to as **1-ads** hereafter.

3.2. Spectroscopic Properties of PVG-Adsorbed 1. Figure 3 shows normalized UV-visible spectra of **1-ads** (in air) and **1** (in H₂O). The spectra are very similar. The λ_{\max} value of **1-ads** in air is 419 nm. The λ_{\max} value of **1** in H₂O is 422 nm. There appears to be only a slight decrease of the 500–600 nm absorption of heterogeneous **1** compared to homogeneous **1**. The absorption of **1-ads** consists of a band in the visible region between 500 and 700 nm, indicating that it may be excited upon the absorption of light in this range. The similarity between the two spectra suggests the **1-ads** may have similar properties to serve as a ¹O₂ photosensitizer.

3.3. Detection of Singlet Oxygen. In D₂O, unlike H₂O, ¹O₂ luminescence is easily detected at 1270 nm from the 532 nm irradiation of **1-ads** (Figure 4). The generation of ¹O₂ is based on a bimolecular sensitization reaction, represented by the

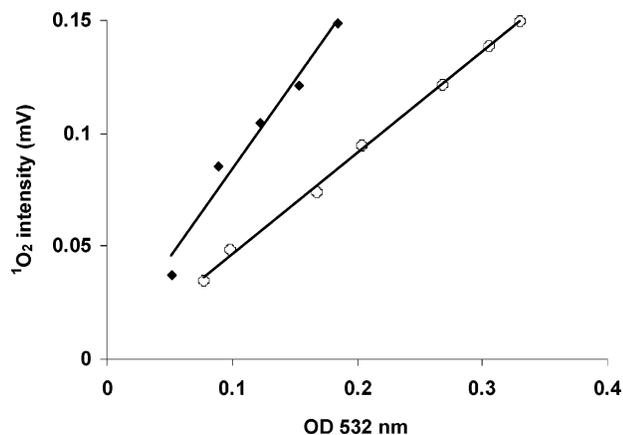


Figure 5. Time-resolved quantum yield measurement. $^1\text{O}_2$ emission intensity as a function of absorbance in D_2O with an excitation wavelength of 532 nm. The solid squares represent *meso*-tetra(4-sulfonatophenyl)porphine dihydrochloride with $\Phi_{\Delta} = 0.63$ in D_2O . The open circles represent free **1** measured to be $\Phi_{\Delta} = 0.43 \pm 0.07$ in D_2O .

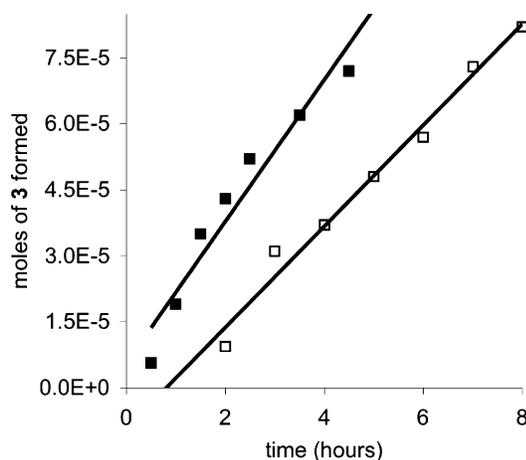
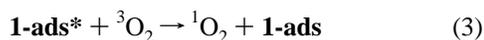


Figure 6. Photooxidation of *trans*-2-methyl-2-pentenoate anion by **1-ads** as a function of irradiation time in D_2O [solid squares ($y = 2 \times 10^{-5}x + 6 \times 10^{-6}$; $r^2 = 0.9394$)] and H_2O [hollow squares ($y = 1 \times 10^{-5}x + 9 \times 10^{-6}$; $r^2 = 0.9832$)].

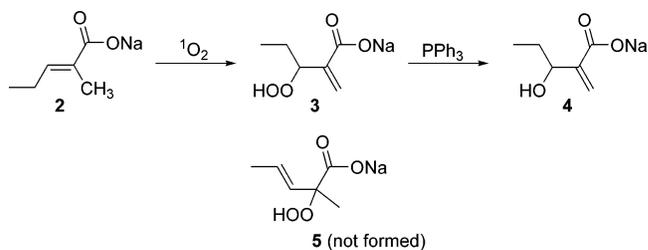
reaction between the electronically excited sensitizer, **1-ads**^{*}, and the acceptor, $^3\text{O}_2$. It is possible that **1-ads**^{*} is quenched by O_2 which is adsorbed itself or, simply, O_2 collides with excited **1-ads**. The formation of $^1\text{O}_2$ is predominantly a triplet quenching process (eq 3).



The 1270 nm luminescence decay is exponential, and the lifetime of $^1\text{O}_2$ in D_2O is found to be 65 μs , which matches the literature value.⁴¹ After $^1\text{O}_2$ is generated, its diffusion into the surrounding solution can take place. Ogilby et al.⁴ suggested that if one assumes a typical diffusion coefficient for oxygen,⁴² D , in liquid of $\sim 3 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$, then the distance traveled by $^1\text{O}_2$ is approximately 62 μm in D_2O . The difficulty in detecting $^1\text{O}_2$ in H_2O arises from its short lifetime (3.1 μs)⁴³ due to efficient electronic to vibronic energy transfer between the two.⁴⁴

Figure 5 shows $^1\text{O}_2$ emission intensity over a range of absorbances for **1** in homogeneous D_2O solution. The literature value of Φ_{Δ} for the reference sensitizer *meso*-tetra(4-sulfonatophenyl)porphine dihydrochloride is 0.63 in D_2O .⁴⁵ Thus, Φ_{Δ} for the formation of $^1\text{O}_2$ from **1** is 0.43 ± 0.07 in D_2O , using *meso*-tetra(4-sulfonatophenyl)porphine dihydrochloride as

SCHEME 2



a reference. Triplet–triplet annihilation appears to be negligible at absorbances ranging from 0.03 to 0.60 from excitation at 532 nm, as indicated by the $^1\text{O}_2$ intensity showing a linear correlation with the absorption of the complexes. Because there is no reference sensitizer–PVG complex, we were not able to determine the Φ_{Δ} value for the formation of $^1\text{O}_2$ with **1-ads**. PVG-adsorbed **1** appears to retain its photosensitization property in aqueous solution. Unlike porous silicon nanocrystals,^{44,46,47} PVG is not found to produce $^1\text{O}_2$ in D_2O in the absence of adsorbed **1**. Furthermore, the 1270 nm luminescence of $^1\text{O}_2$ in H_2O appears at the noise level of the instrument. However, we show that $^1\text{O}_2$ is formed in H_2O by indirect trapping experiments. The results described next show that $^1\text{O}_2$ produced in the **1-ads** photosensitized reaction can react with *trans*-2-methyl-2-pentenoate anion (**2**).

3.4. Photooxidation of *Trans*-2-methyl-2-pentenoate Anion (2**).** Unlike the aerobic UV irradiation of PVG reported to form superoxide,⁴⁸ the visible light irradiation of **1-ads** generates $^1\text{O}_2$, which can diffuse out of the silica matrix, and is then trapped by a *trans*-2-methyl-2-pentenoate anion (**2**) in a surrounding aqueous solution. The **1-ads** sensitized photooxidation of **2** affords one product, hydroperoxide **3** (Scheme 2). The reaction is monitored by NMR. No other products were detected, which would be expected if superoxide were present. Formation of **3** and not regioisomer **5** indicates that the methyl protons are more acidic, in which the resulting double bond is conjugated with the carboxylate group. Hydroperoxide **3** reacted with triphenylphosphine and converted to alcohol **4**, also characterized by NMR. The yield of **3** was dependent on whether the **1-ads** sensitized photooxidation of **2** was conducted in D_2O or H_2O . Figure 6 shows that this $^1\text{O}_2$ “ene” reaction is found to be about 2 times faster in D_2O than H_2O . Hydroperoxide **3** was formed more rapidly in D_2O than H_2O (cf. 16.9 to 3.9%) after 2 h. In H_2O , photooxygenation of **2** with **1-ads** formed **3** in 32.8% yield after 8 h. The results support the conclusion that the reaction of $^1\text{O}_2$ with **3** occurs in the surrounding H_2O or D_2O solution. Previous data led to a similar conclusion that alkene oxidation is more efficient in D_2O than H_2O .³⁵ The results establish the feasibility of carrying out the heterogeneous sensitization process with **1-ads** in which $^1\text{O}_2$ is released into surrounding D_2O and H_2O .

4. Conclusion

PVG is a cation-binding solid. We took advantage of this binding property to examine PVG as a support for the cationic photosensitizer **1**. Sensitizer **1** adsorbs to PVG and gives a stable material, which does not dissociate in water at room temperature. Singlet oxygen is generated in the surrounding aqueous solution upon irradiation of the adsorbed complex. The excited state of **1-ads** is quenched by O_2 to give $^1\text{O}_2$, which can be detected in the surrounding aqueous solution. The heterogeneous system described in this paper could have application in ridding wastewater of *E. coli*.

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